



Resistenzen  
bei Tier und Mensch -  
gemeinsame Forschung in Deutschland



# Significance of environmental contaminations on the development of bacterial resistance against antibacterial agents in indicator animals

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## Topics

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### RESET I



**Incorporation of antimicrobial substances in vegetables from manure-fertilized soil and microbiological effects**

**Significance of dosage and treatment duration on the development of bacterial resistance against antibacterial agents in farm animals**

### RESET II

- 1. Impact of antibiotics in plants on bacterial susceptibility *in vitro* and *in vivo***
- 2. Influence of different oral dosage forms on environmental pollution and bacterial susceptibility**

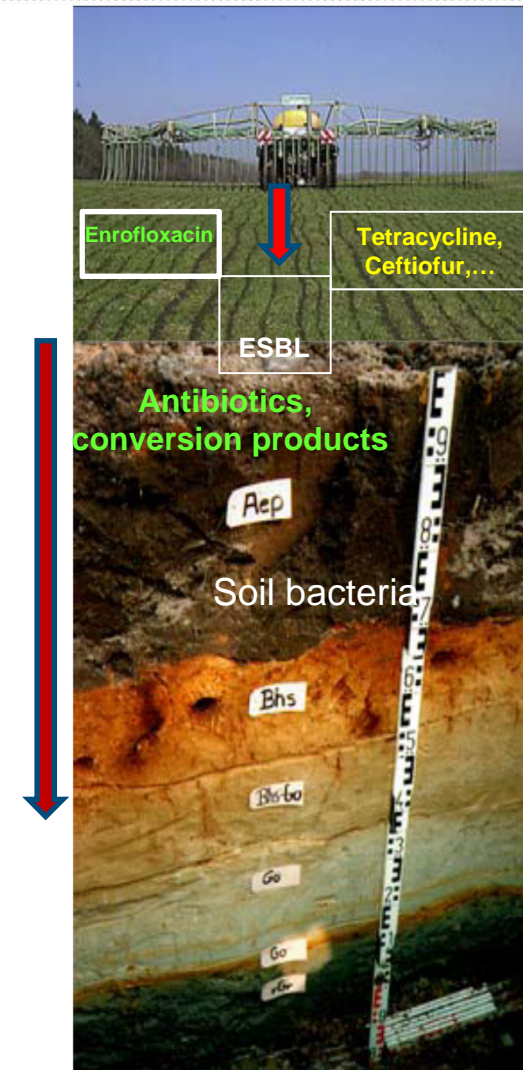
## Background I: first funding period

### Tracing the pathways/exposure routes of veterinary antibiotics in agricultural systems

...from the source (animal production – excrements) - via fertilization of soil by liquid manure - transfer into crop plants ?

- Model studies under farming conditions (approach to agricultural practise)

- Hydroponic model systems



# Seeding of white cabbage and leeks



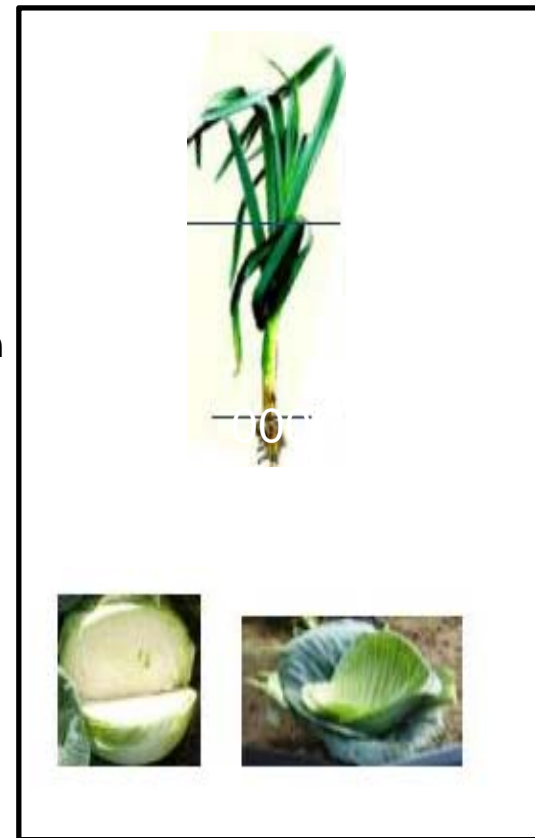
seedlings



14 days after planting: Fertilization in grooves next to the crop row



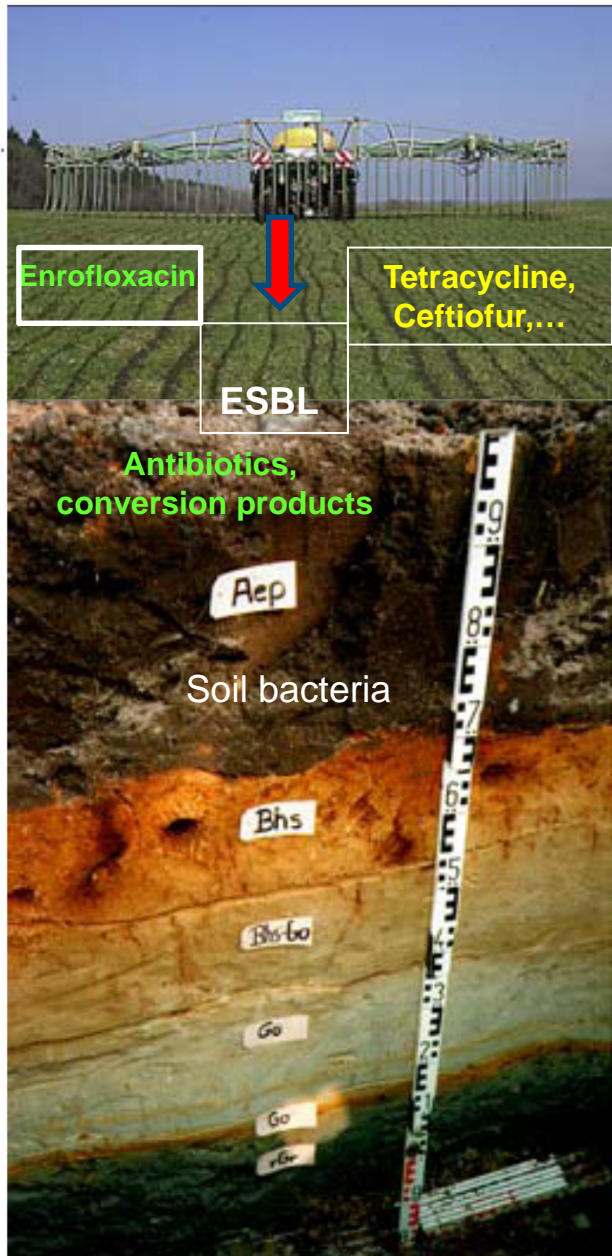
Cabbage field after harvesting: Sampling in Z-configuration



**Trace-Analysis: Antibiotic residues, microbiology**



## Results



**Slurry dispersion on fields:**

**Impact of antibiotics and bacteria into soil**

➔ **ESBL- *E. coli* survive in soil and on plants until harvest (4-5 months)**

➔ **Transfer of antibiotics from soil into edible parts of vegetable:**

**Enrofloxacin, Tetracycline,..**

**Effects of low AB-concentrations in food and feed? Risk for consumers?**

(Acc. to G.Hamscher, 2004)

# Antibiotic contaminated food plants - a risk to consumers?

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## Comment of the Federal Institute for Risk Assessment (BfR)\*

⇒ "... that on the basis of the available data to the amount of AB residues (...) in food plants is from a toxicological point of view no risk for the consumer to be expected."

⇒ "... the risk of resistance development in bacteria, which may be of importance for the humans, by long-term exposure to small amounts of antibiotics on plant foods .... about uptake and retention... in plant feed and food ....is conclusively not assessable."

**→ Combined effects by incorporated antibiotics and microbial contamination?**

\* Release No. 019/2010 of 15 BfR February 2010

# Topics

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## RESET I



Incorporation of antimicrobial substances in vegetables from manure-fertilized soil and microbiological effects

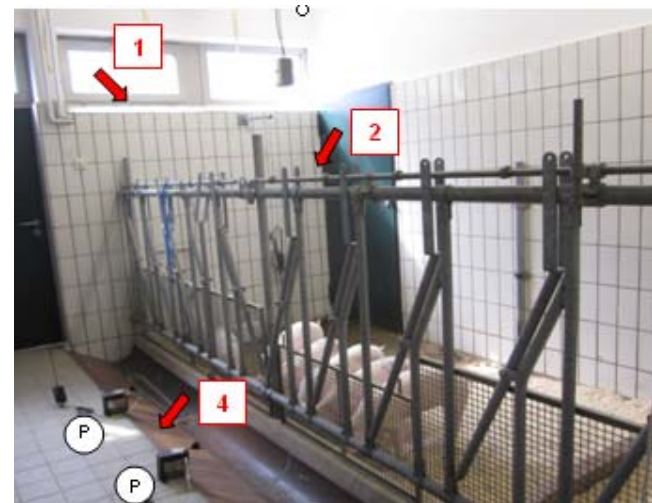
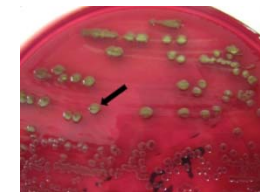
Significance of dosage and treatment duration on the development of bacterial resistance against antibacterial agents in farm animals

## RESET II

- 1. Impact of antibiotics in plants on bacterial susceptibility *in vitro* and *in vivo***
2. Influence of different oral dosage forms on environmental pollution and bacterial susceptibility

## Background II: first funding period

- Studies on the susceptibility of intestinal *E. coli* after oral (enrofloxacin) and parenteral (ceftiofur) antibiotic treatment in poultry and pigs
- Bioavailability determination of antibiotics (plasma)
- Determination of environmental antibiotic burden after treatment (dust and aerosol)



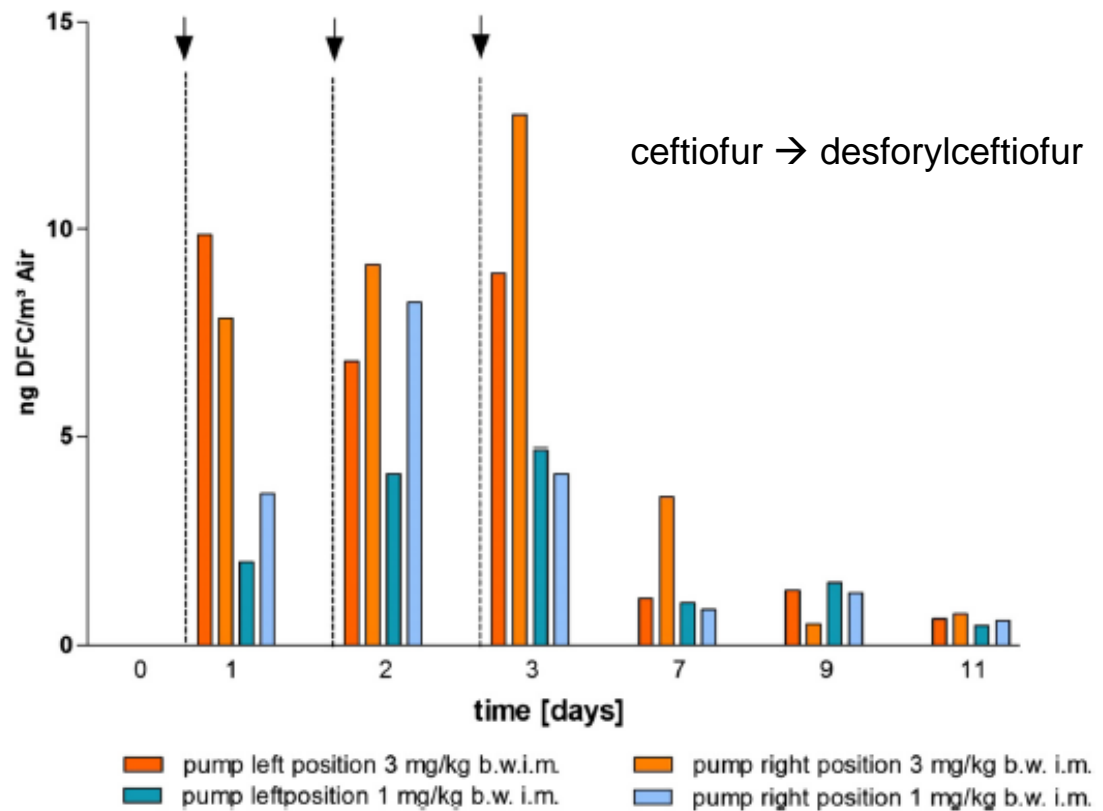
Scherz 2013, Zessel 2012



## Environmental contamination after parenteral administration of ceftiofur

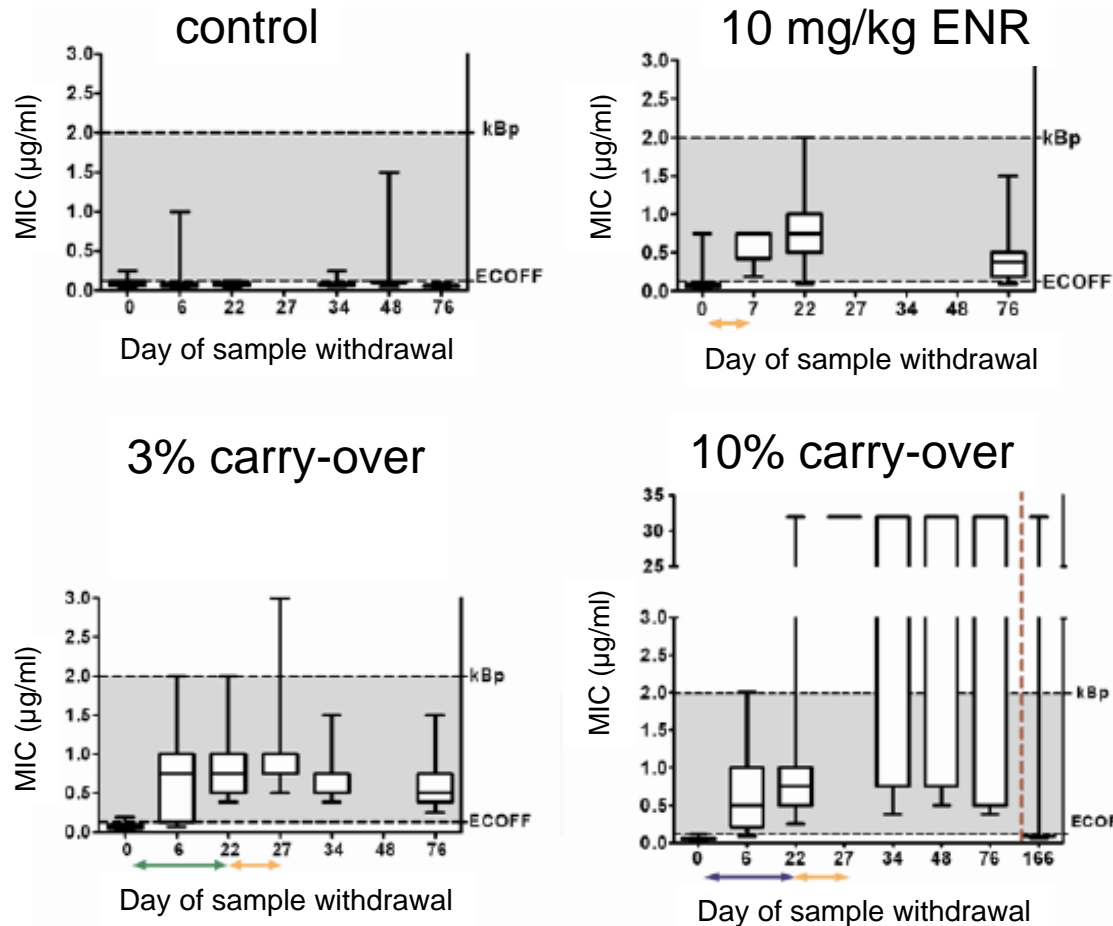
**Antibiotic residues in the stable after antibiotic treatment with enrofloxacin (poultry, oral administration) and ceftiofur (pigs, parenteral administration) correlate with drug dosage**

→ is a high environmental pollution associated with shifts in bacterial susceptibility of untreated animals?



Beyer et al. 2015

# Influence of subtherapeutic ENR-dosages on bacterial resistance of commensal *E. coli* in poultry



**Subtherapeutic dosages of enrofloxacin result in MIC-increases in commensal intestinal *E. coli***

→ Cabbage feeding  
→ MIC-shifts?

## Topics

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1. Impact of antibiotics in plants on bacterial susceptibility *in vitro* and *in vivo*
2. **Influence of different oral dosage forms on environmental pollution and bacterial susceptibility**

Enrofloxacin (ENR)

## RESET II: Topics

### Part 1. Antibiotics in plants and their impact on bacterial resistance



- *in vitro*
- *in vivo*

### Part 2. Influence of different oral dosage forms on environmental pollution and bacterial susceptibility

→ powder



→ pellets



→ granulate

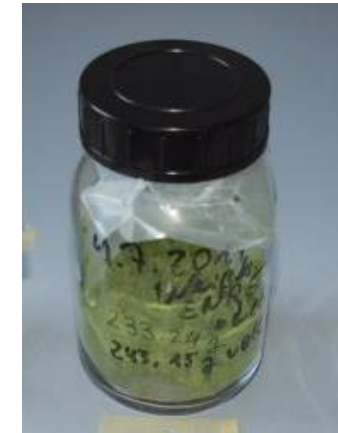




# Cabbage processing



- collection of leaves
- fresh leaf material
- bacterial inhibition tests
- freeze drying
- grinding



- ENR determination (LC-MS/MS)
- bacterial inhibition tests
- resistance development (MIC shift)
- animal feeding experiments in mice and poultry

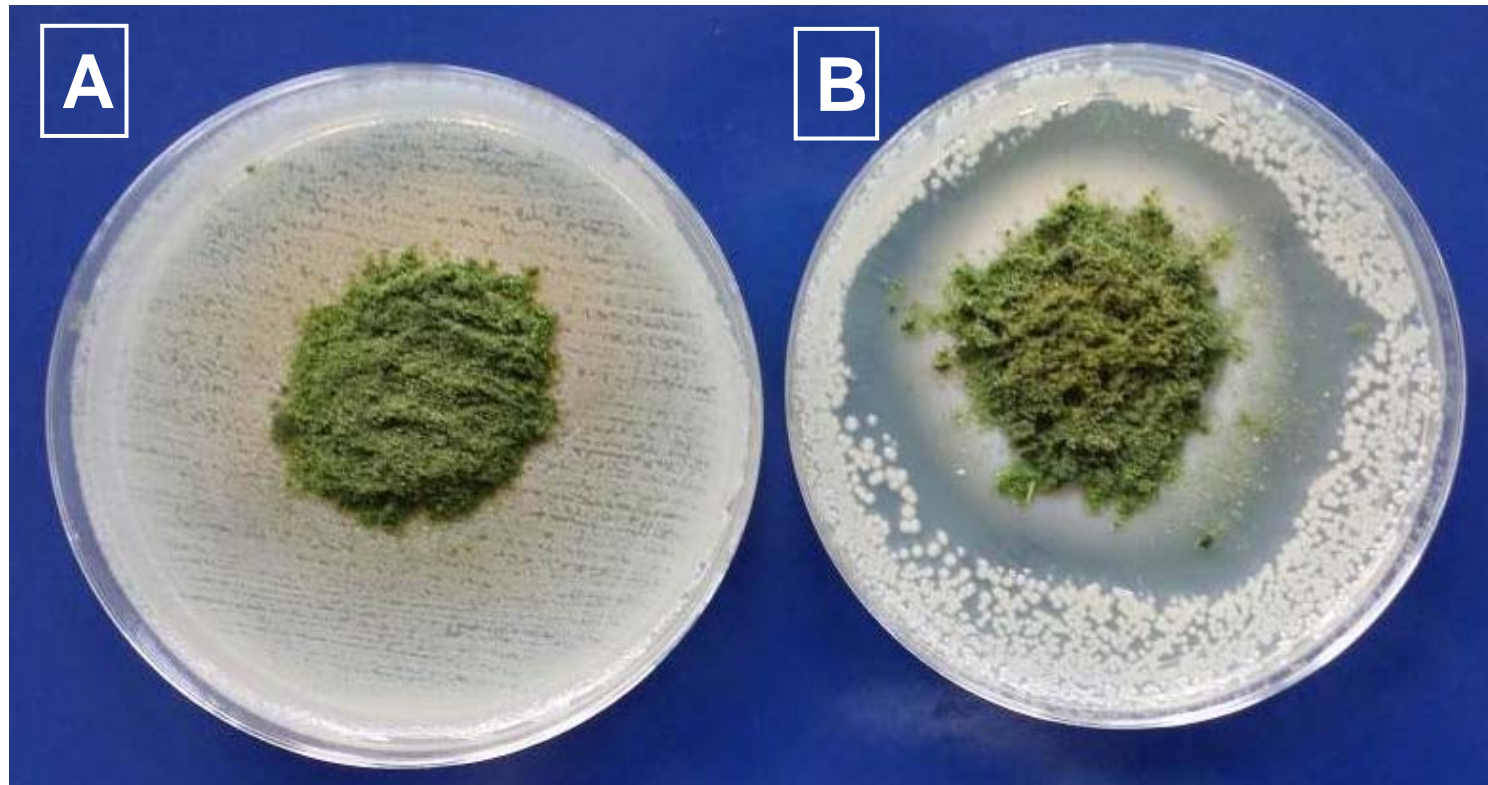


collection of xylem sap



- ENR determination (LC-MS/MS)
- bacterial inhibition tests

## ENR release out of plant material?

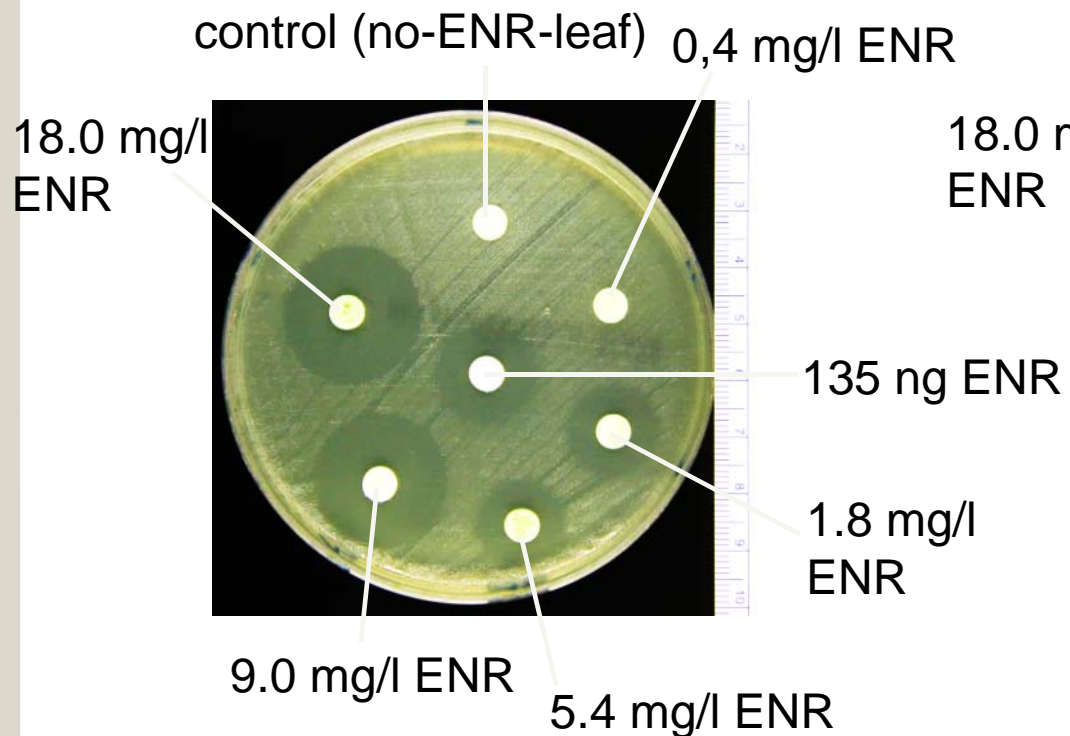


- A:** Cabbage without ENR on Mueller-Hinton-agar (*E.coli* (MIC: 0.015 mg/l))  
**B:** Cabbage with ENR (58 µg/g) on Mueller-Hinton-agar (*E.coli* (MIC: 0.015 mg/l))

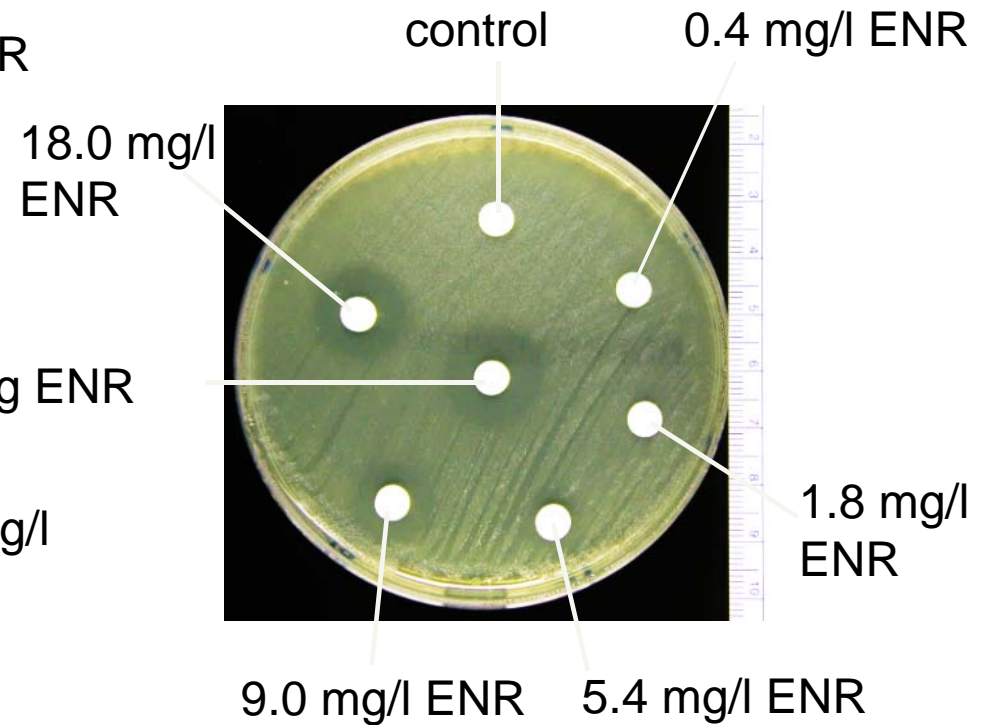
## Leaf extracts and xylem sap inhibit *E. coli* growth

- bacterial inhibition test on nutrient rich agar plates
- plating of *E. coli* culture and addition of cabbage extracts

### leaf extracts: 15 µl/filter disc



### xylem sap: 15 µl/filter disc

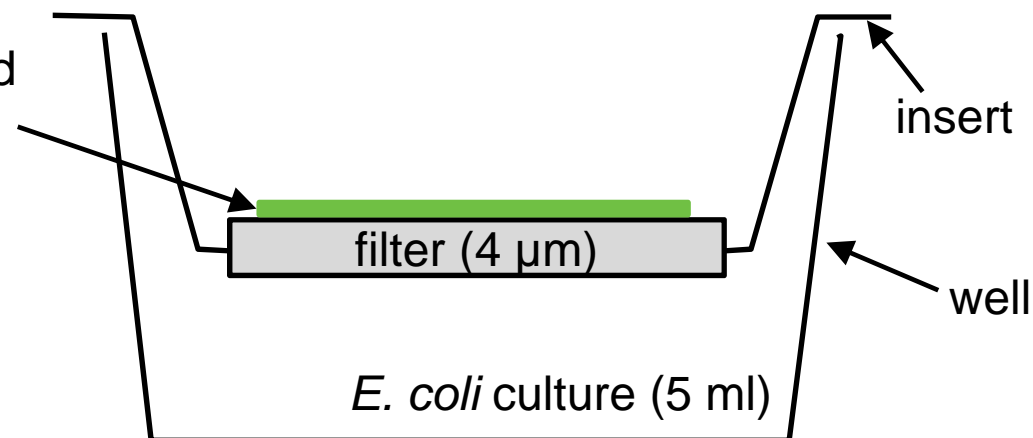


## Incubation of *E. coli* with ENR-containing cabbage

### ➤ bacterial resistance development?

cabbage:

- freeze dried + ground
- 155 µg/g ENR
- 1 g per insert

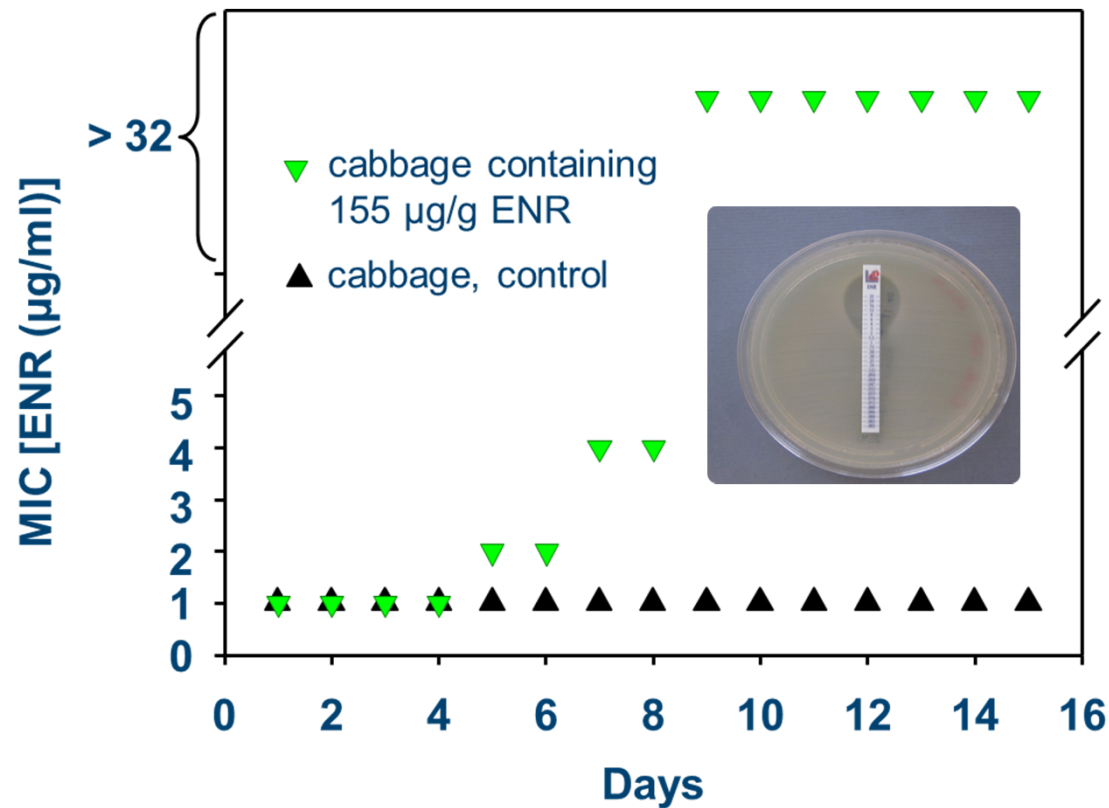


15 days: every 24 h withdrawal of an aliquot from the *E. coli* culture

MIC shift assay: plating on agar plates containing increasing concentrations of ENR and Etest®



## Incubation of *E. coli* with enrofloxacin-containing cabbage leads to a shift in the minimum inhibitory concentration (MIC) of ENR



1g of cabbage with 155 µg/g ENR, incubation with *E. coli* culture  
 MIC shift assay: plating on agar plates containing increasing concentrations of ENR and Etest®

## Animal feeding experiments I

### Mice

- feeding of ENR containing cabbage, 0.1 mg ENR/kg b.w. over 21 days followed by 5 days therapeutic dosage
- Samples of faeces and urine (cage)



### Results

- detection of ENR in urine of mice in amounts of 0.4-2.7 ng/ml urine
- no MIC shift in *E. coli* isolates from mice faeces, only wild type isolates
- no development of resistance in commensal *E. coli* of mice (MIC: 0.032 - 0.064 µg/ml)

## Animal feeding experiments II

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### Chicken

- ENR over 21 days in subtherapeutic concentration, then ENR in therapeutic dosage for 5 days
- Samples of faeces and blood at several days
- Establishment of HPLC-method to determine ENR in faeces and blood samples of chicken

### Results

- MIC of commensal intestinal *E.coli* → first MIC shift on day 6 in the cabbage group
- Content of ENR in faeces and blood → analysis is still in progress

## Summary part 1



Poster: Langenkaemper et al.

- **cabbage has a considerable capacity for uptake of ENR in hydroponic culture**
- **ENR that went through the chain -cabbage uptake, freeze drying, and grinding- is still active as an antibiotic substance (inhibition effect on growth of *E. coli* culture, MIC shift)**
- **mice receiving low doses of ENR via cabbage did not develop resistant commensal *E. coli*, ENR was detected in mice urine**
- **in chicken receiving low doses of ENR via cabbage a MIC-shift was detectable, analysis of bioavailability is after establishment of the determination method in progress now**



## RESET II: Topics

### Part 1. Antibiotics in plants and their impact on bacterial resistance



- *in vitro*
- *in vivo*

### Part 2. Influence of different oral dosage forms on environmental pollution and bacterial susceptibility

→ powder



→ pellets



→ granulate



## Aims part 2

### Influence of different oral dosage forms with antibiotics...

1. ... on the susceptibility of commensal *E. coli* in treated pigs
2. ... on the carry over in the environment
3. ... on the development of resistant *E. coli* in non treated pigs kept in the same stable



→ powder



→ pellets



→ granulate





**treated group**

**control group**

1

2

(P)

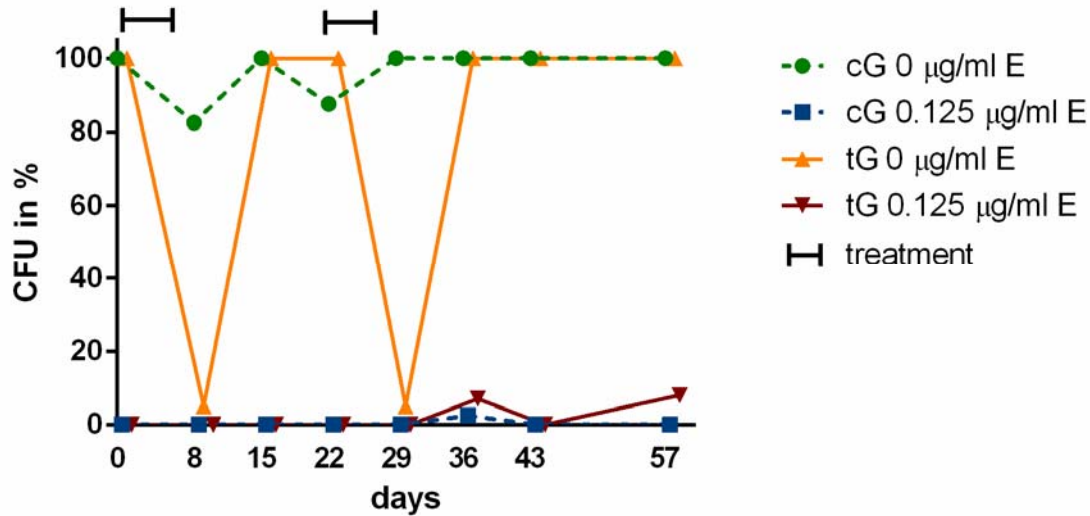
(P)

(X) sedimentation dust  
(P) air sampling pump



# Bacterial susceptibility

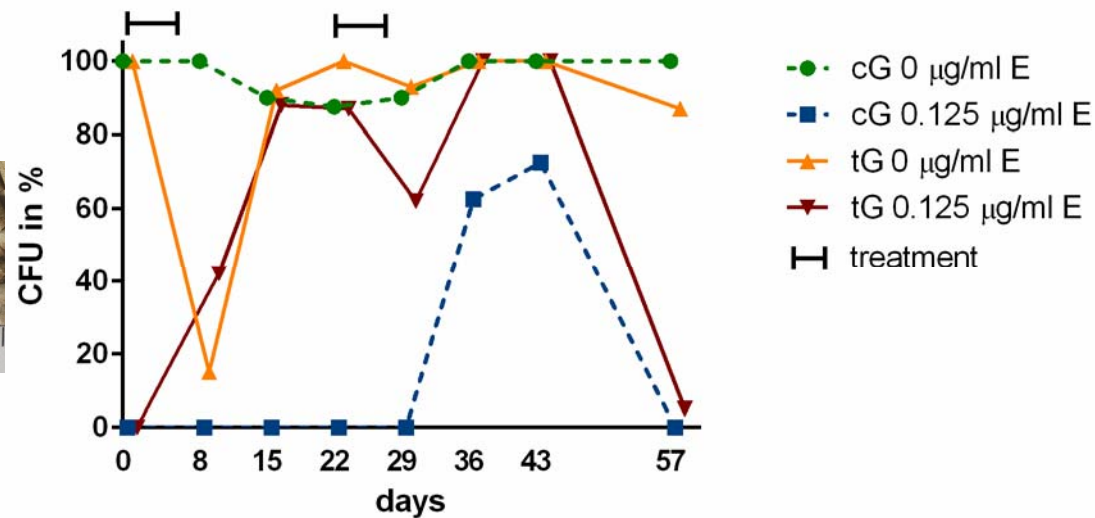
powder



cG = control group (sentinel)  
tG = treated group

CFU= colony forming units  
E = enrofloxacin  
—|— = treatment period

pellet



Diss. 2017; Hagedorn

## Results and summary part 2

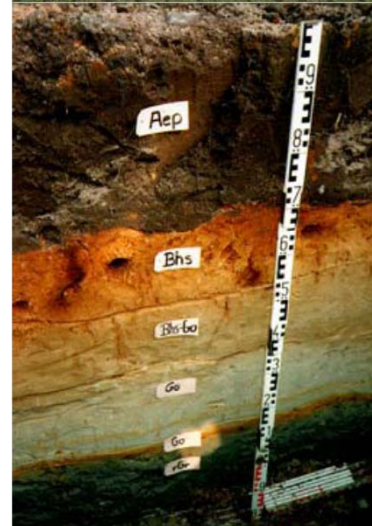


- **similar oral bioavailability of ENR in all dosage forms**
- **greatest MIC shift during pellet feeding, followed by granulate feeding (pellet > granulate >> powder)**
- **higher values of enro- and ciprofloxacin in sedimentation dust during oral treatment via powder and granulate**
- **no correlation between dust pollution and development of bacterial resistance**
- **MIC shift in sentinels detectable (granulate & pellet)**



# Results & Outlook

- Risk for untreated animals
- Environmental pollution
- Different dosage forms?



**New strategies?!?**

**Consumers**

# Thanks...

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**Frauke Mellis**  
**Marie-Thérèse Hanneforth**

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**Clinic for poultry, TiHo Hannover**

**Institute of Particle Technology, TU Braunschweig**

**All RESET-members...for the excellent collaboration in the last years!**



collaborative research project

<http://www.reset-verbund.de>

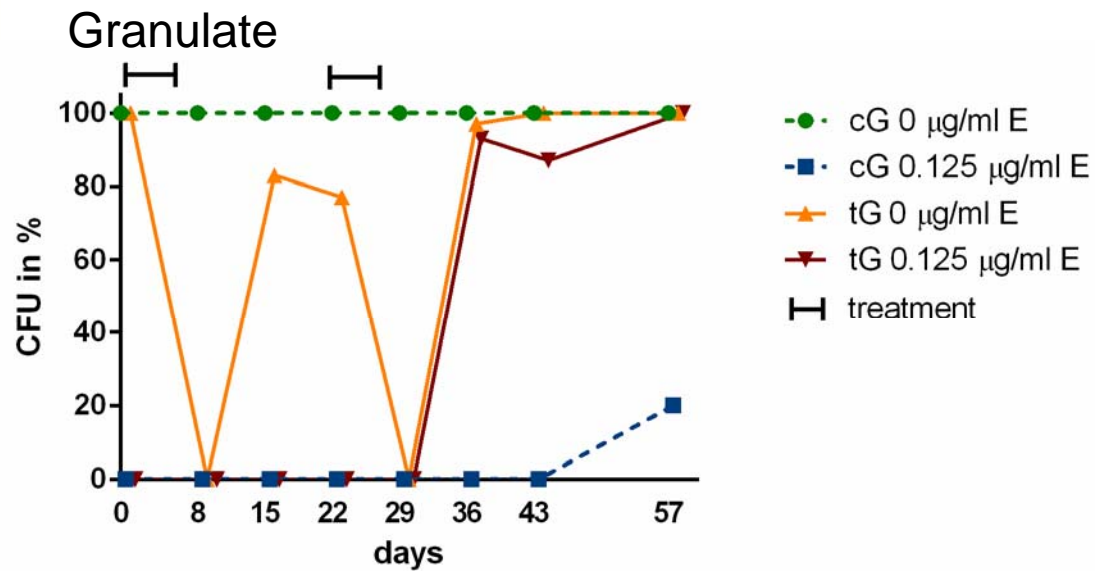
ESBL and (fluoro)quinolone  
Resistance in Enterobacteriaceae  
Project No. 01KI1313A

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## MRI

Husam Aroud  
Christine Schwake-Anduschus  
Annette Meyer-Wieneke  
Dagmar Oeldemann  
Beate Lippert  
Frauke Mellis  
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## TiHo Hannover

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Stefanie Mielke-Kuschow  
Manfred Kietzmann

## Paderborn University

Manfred Grote



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## Results

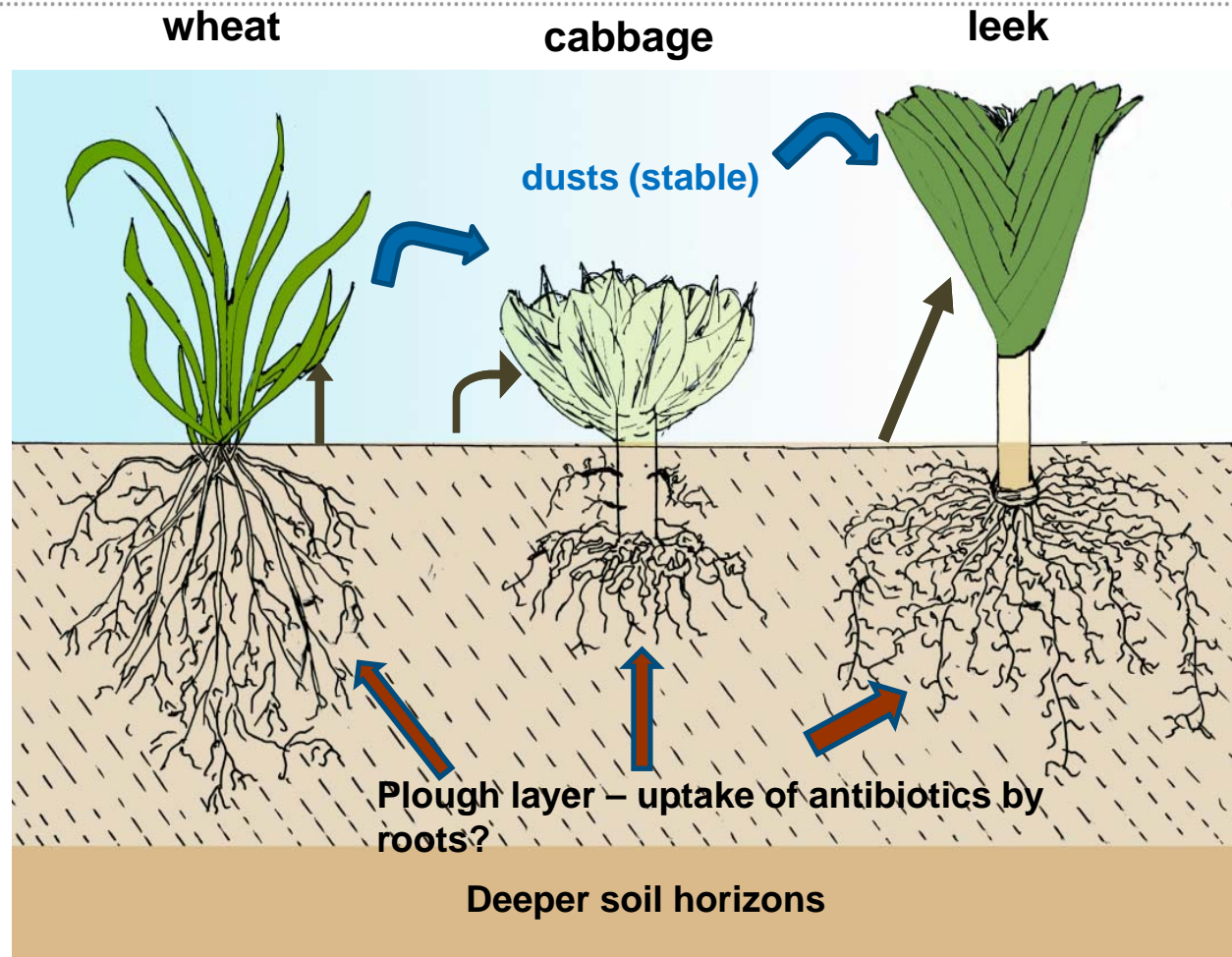
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**ESBL- *E. coli* survive in soil and on plants until harvest (4-5 months)**

**Transfer of antibiotics from soil into edible parts of vegetable:  
Enrofloxacin, Tetracycline,...**

**Effects of low AB-concentrations in food and feed? Risk for consumers?**

## Possible pathways of antibiotics (and bacteria) to plants....



....exposure routes of antibiotics into the food chain

..... continuing uptake of **low concentrations of antibiotics**  
by contaminated crop plants:  
→Promotion of **initiation of resistance?**

Possible exposure of consumers to antibiotics by **contaminated crop plants:**  
→a risk to consumers?

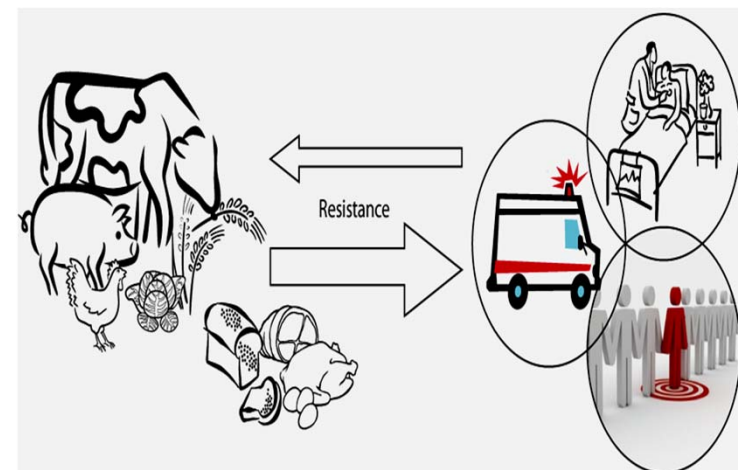
\*sub-inhibitory concentration of  
antibiotics:  
below the *minimum inhibitory  
concentration* (MIC)

## Outlook

\*In future research must be considered:

Collateral effects of sub-inhibitoric doses  
of microbial active contaminants  
in **food** and **feed**,

such as effects on the **intestinal microbiome**  
in **man** and **animal**.



\*According to: T. Looft, T. A. Johnson, H. K. Allen et al.,  
*In-feed antibiotic effects on the swine intestinal microbiome*, PNAS, January  
31, **2012**, 109, 1691 - 1696

# Experimentelle Ergebnisse #

*(Zum Vergleich der Ergebnisse: Folie von L. Kreienbrock, Allgemeiner Vortrag)*

Behandlung von Tiergruppen mit Antibiotika führt zu Verschleppung in die direkte Tierumgebung

Gemüsepflanzen (Weißkohl, Porree) nehmen antimikrobiell wirksame Stoffe aus Böden auf

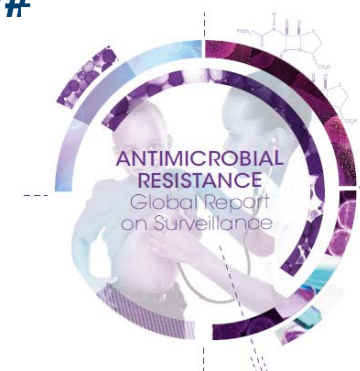
ESBL-*E. coli*, die zur Gülle dotiert worden sind, lassen sich in Boden und Gemüse nachweisen

Die Exposition mit subtherapeutischen AB- Konzentrationen übt einen Selektionsdruck aus



- Antibiotic resistance is one of the biggest threats to global health today.  
It can affect anyone, of any age, in any country
- Antibiotic resistance occurs naturally, but misuse of antibiotics in humans and animals is accelerating the process
- A growing number of infections are becoming harder to treat as the antibiotics used to treat them become less effective
- Antibiotic resistance leads to longer hospital stays, higher medical costs and increased mortality
- A post-antibiotic era—in which common infections and minor injuries can kill— is a very real possibility for the 21<sup>st</sup> century#

**Antimicrobial resistance: global report on surveillance.  
World Health Organization, 2014**



## Fluorquinolones

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- **broad-spectrum antibiotic drugs, gyrase and topoisomerase inhibition in bacterial species, prevents bacterial cell division**
- **WHO list of critically important antimicrobials<sup>1</sup>**
  - **- reserve antibiotics**
  - **- veterinary use is only considered acceptable provided that no alternative treatment is available**

<sup>1</sup>**Answer to the Request for scientific advice on the impact on public health and animal health of the use of antibiotics in animals**, Antimicrobial Advice ad hoc Expert Group European Medicines Agency, 2014

## Bacterial resistance against fluoroquinolones

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- mutations of gyrase and topoisomerase genes
- efflux mechanisms
- plasmid encoded resistance
  
- Sub-inhibitory levels of antibiotic substances

accelerate the emergence and spread of antibiotic-resistant bacteria among humans and animals<sup>2</sup>

bacterial resistance development at antibiotic concentrations several 100-fold below sub-minimal inhibitory concentrations, e.g. for ciprofloxacin<sup>3</sup>

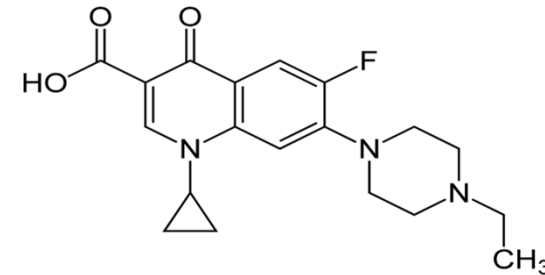
<sup>1</sup>Sköld 2011 *Antibiotics and antibiotic resistance*. Wiley

<sup>2</sup>Andersson & Hughes 2014 *Nature Reviews Microbiology* **12**, 465

<sup>3</sup>Sandegren 2014 *Upsala J. Med. Sci.* **119**, 103

## Enrofloxacin

- most frequently used fluoroquinolone in animal production
- biotransformation, i.e. removal of ethyl group: ciprofloxacin<sup>1</sup>
- environmental distribution: is found in animal urine and excrements, waste water, is stable in slurry and soil<sup>1</sup>
- can be taken up by plants such as cucumber<sup>2</sup>, lettuce<sup>2</sup> and red cabbage<sup>3</sup> from the soil ( $\mu\text{g}/\text{kg}$  f.w. range)
- has been detected in supermarket vegetables in China: detection frequency of quinolones >90%, concentrations from 10 to 193  $\mu\text{g}/\text{kg}$  d.w.<sup>4</sup>
- leek, white and red cabbage readily take up ENR in hydroponic cultures (up to 6 mg/kg f.w. in leaves)<sup>3, 5</sup>



<sup>1</sup>Rusu et al. 2015 Environ. Chem. Let. **13**, 21  
Agron. Res. **8**, 807

| <sup>2</sup>Lillenberg et al. 2010

<sup>3</sup>Chowdhury et al. 2016 J. Verbrauch. Lebensm. **11**, 61 | <sup>4</sup>Wu et al. 2011 Huan. Kexue/Environm. Sci. **32**, 1703

<sup>5</sup>Grote et al. 2009 J. Verbrauch. Lebensm. **4**, 287

# Experimental model

- **substance:** enrofloxacin
- **animal:** pig  
→ treated group and control group
- **indicator bacteria:** commensal intestinal *E. coli*  
→ ECOFF\* for enrofloxacin:  $< 0.125 \mu\text{g/ml}$
- **treatment:** via different oral dosage forms



\*ECOFF: epidemiological cut-off